Original Article

**Study on Some Ectoparasitic Diseases of Catfish, *Clarias gariepinus* with their Control by Ginger, *Zingiber officiale***

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**Abstract**

Parasitological investigation was performed in one hundred naturally collected Nile catfish *Clarias gariepinus*. It revealed skin and gill infestation with ectoparasitic protozoan *Trichodina* and *Epistylis* and monogenean *Gyrodactylus spp.* with average 20, 15 and 25%, respectively. The average of the total ectoparasitic infestation in the examined catfish was 60%. The significance of herbal control measures using ginger *Zingiber officiale* as a new method to eradicate such parasites, was evaluated. A bioassay of 96 hours LC_{50} of ginger emphasized that its value was 192mg/L. The herbal control with ginger was safe and effective to treat the ectoparasitic protozoa *Trichodina* and *Epistylis spp.* at dose 20mg/L, but not suitable for treating the monogenean *Gyrodactylus spp.*. The estimation of dissolved oxygen, pH and total ammonia of aquarium water pre and during treatment was indicated that the use of ginger in aquaculture operation will improve its water quality especially the total ammonia.

**Key Words:** *Clarias gariepinus*, *Trichodina spp.*, *Epistylis spp.*, *Gyrodactylus spp.*, Ginger, Aquaculture.

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**Introduction**

The African catfish, *Clarias gariepinus* is one of the most important fish species for an economically successful aquaculture. However, farmers are constraint with massive fry and fingerling mortalities, especially in culture system due to the invasion of parasites. The protozoan *Trichodina spp.* and the monogenean *Gyrodactylus spp.* were the dominant parasites

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which were aggravated pathological effects on fish and increasing mortality (Akoll, 2005). Moreover, Kristmundsson et al. (2006) stated that the rearing environment with a subsequent increase in biomass and decreasing temperature caused a proliferation of trichodinids. Also, during parasitological surveys in Chogqing, China, Zhao and Tang (2007) investigated three trichodinids infesting a freshwater fish and a mollusc. Hubert and Warner (1975) recorded that *Epistylis* first appeared on the tips of both dorsal and pectoral fin spines. When untreated, *Epistylis* colonies spread and eventually covered much of the anterior region of the body and causes mortality of catfish. Paperna (1996) mentioned that fish heavily infested with *Gyrodactylus* appeared pale, due to excessive mucus secretion and epithelial proliferation. In the more heavily infested skin zones, there are skin erosion, desquamation of the skin epithelium, focal hemorrhagic lesions and severe mortalities. In addition, Boeger et al. (2005) reported that viviparous gyrodactylids exhibit extraordinary species diversity and broad host range if compared to other monogenean groups.

In the last few years, the herbal treatments were developed for replacing the chemical used for eradication of the most harmful parasites infesting terrestrial animals. In this respect, several herbs were used in pest and parasitic control as ginger which contains over four hundred chemically active ingredients, has long been used for treatment a variable diseases. It has a wide range of actions against parasites (Leo, 2001). In addition, Pro (2003) stated that ginger is the newest homeopathic treatment to be suggested. Practically, Colorni (1987) succeeded to treat fish infected with Cryptocaryon irritans with ginger for ten days with changing water every three days.

The present study was planned to endeavour of evaluating the possible use of new herbal control (Ginger) against some ectoparasitic protozoa and monogenea that infested *Clarias gariepinus* fish with calculating optimum dose that can completely eradicate these parasites by determination of the lethal concentration 50 of ginger and safety margin for such fish.

**Materials and Methods**

**Parasitological examination of fish:**
A total of 145 freshwater *Clarias gariepinus* ranged from 200-250g body weight were collected alive from El-Monib, Giza governorate fish market and transported immediately to the lab in large plastic bags partially filled with water and containing air according to Langdon and Jones (2002). The fish were examined for gross clinical signs and pathological lesion. The necropsy technique of parasitological examination of skin, fins and gills was carried out for the presence of external parasites with recording the average and intensity of the isolated parasites from one hundred catfish, the other 45 catfish were kept for applying the treatment. The recovered protozoa and monogenea were fixed, preserved and stained according to Paperna (1996), Langdon and Jones (2002).

**Ginger (Zingiber officiale):**
It was obtained as patent preparation in a Tablet form from MEPACO. Ginger is belonging to the Zingiberaceae plant family. Ginger contains several chemical components as outlined by Murray (1995): starch (50%), protein (9%), lipids (including glycerides, phosphatidic acid, lecithins, and fatty acids, 6-8%), protease (2.26%) and volatile oils (including gingerol, shogoal, zingiberene and zingiberol, 1-3%). In addition, it contains vitamins A and B3 (niacin)

**Ginger lethal concentration-50 (LC$_{50}$):**
A total number of 100 fish were divided into ten groups. The fish were exposed to different concentrations of ginger (Table 2), kept under observation for 96 hours and numbers of dead fish were recorded. The results were statistically analyzed according to Behrens and Karber (1953) by this equation:

$$AXB\quad LC_{50} = \frac{\text{Largest dose} - \sum}{N}$$
Where A is the mean of dead fish between two successive doses.
B is dose differences between two successive doses.
N is the total number of fish.

**Efficacy of ginger bath on the ectoparasites infesting *C. gariepinus*:**
A total of 45 alive naturally infested *C. gariepinus* were divided in three groups of 15 fish each in a volume of water (100x 40x 15 cm, 60 liters) with good aerated conditions. The safe level of ginger applied to infested fish was the 10 times lower than the value of ginger 96 hours LC$_{50}$ (Boyd 1998). It was approximately 20mg/L. The fish in the first group was exposed to 10mg/L ginger while the second group was exposed to 20mg/L ginger and the third group was considered a control.

**The water quality of fish aquarium before and during treatment:**
The fish were kept in the same volume of cleaned water (60-L for 15 fish) with good aeration and dechlorinated tap water, for at least one week before treatment. Water samples from each aquarium were analyzed at room temperature before and during treatment for estimating the dissolved oxygen, pH and total ammonia (Masser and Jensen, 1991). The dissolved oxygen was measured by Cole Parmer 5946-50-oxygen meter, Analog field kit. The pH of water was measured by pH meter (Economy pH/mV/°C, Benchtop Meter) and the total ammonia was measured by the colorimeter apparatus (Model 6030-operating manual-Janway).

**Results**

**Gross macroscopic examination:**
The gross examination of infested fish with the isolated ectoparasitic protozoan, *Trichodina*, *Epistylis* and monogenean *Gyrodactylus spp.* revealed pale skin and excessive mucus secretion. Skin erosion with local hemorrhagic lesions was demonstrated in some infested fish (Fig. 1a&b).

**Parasitological examination:**
The parasitological examination revealed single and mixed infestation of some *C. gariepinus* with the ciliated ectoparasites *Trichodina* and *Epistylis spp.* in addition to monogenean ectoparasites *Gyrodactylus spp.* (Table 1).

Based on the morphological examination of the isolated ectoparasites, they were identified as the following:

**Ectoparasitic protozoa:**
- Phylum: Ciliophora
- Class: Ciliatea
- Subclass: Peritricha
- Order: Mobilina
- Family: Trichodinidae
- Genus: *Trichodina*
- Species: *Trichodina* species
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<table>
<thead>
<tr>
<th>Parasites</th>
<th>No of Examined fish</th>
<th>Total infested fish %</th>
<th>Single infestation %</th>
<th>Mixed infestation %</th>
<th>Intensity of infestation No./ field (10X)</th>
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<tr>
<td>Trichodina spp.</td>
<td>20</td>
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<td>8-10</td>
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<tr>
<td>Epistylis spp.</td>
<td>60</td>
<td>15</td>
<td>30</td>
<td>15-18</td>
<td></td>
</tr>
<tr>
<td>Gyrodactylus spp.</td>
<td>25</td>
<td></td>
<td>6-8</td>
<td></td>
<td></td>
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</tbody>
</table>

Based on the morphological examination of the isolated ectoparasites, they were identified as the following:

**Ectoparasitic protozoa:**
- Phylum: Ciliophora
- Class: Ciliatea
- Subclass: Peritricha
- Order: Peritrishida
- Suborder: Sessilina
- Phylum: Ciliophora
- Class: Ciliatea
- Subclass: Peritricha
- Order: Peritrishida
- Suborder: Sessilina
- Family: Trichodinidae
- Genus: *Trichodina*
- Species: *Trichodina* species

Trichodina spp. is isolated from skin, fins and gills of infested *C. gariepinus*. It is large with disc shaped body. The adhesive disc is saucer shaped. The parasite is provided with several rows of cilia at the circular periphery and the inner circle of toothed denticles. The macronucleus is horse shoe-shaped and micronucleus is small and difficult to be seen in some specimens (Fig. 2).

Epistylis spp. is isolated from skin and gills of infested catfish. It is a sessile contractile ciliate. Stalk is long and non contractile. Often forms a branched colony. The distal end of the organism is surrounded by rapidly moving cilia which appear as a blur (Fig. 3).

**Fig. 2:** Trichodina spp. isolated from the skin and of C. gariepinus (Stained with Giemsa’s stain) (X400).

**Fig. 3:** Epistylis spp. recovered from the gills of C. gariepinus (wetpreparation) (X100).
**Ectoparasitic monogenea:**
- Phylum: Platyhelminths
- Class: Dactylogyroidea
- Family: Gyrodactylidae
- Genus: *Gyrodactylus*
- Species: *Gyrodactylus* spp.

The monogenetic *Gyrodactylus* is detected in gills and skin of infested *C. gariepinus*. It is flat worm with one pair of projection at its anterior pole. It can be distinguished from other monogeneans by the absence of eye spots and the occurrence of the embryos in the mid-region of the body (Viviparous monogenean). Posterior end has the organ of fixation, opisthaptor, which is guarded with a number of marginal hooklets and a central one pair of hooks (Fig. 4, 5).

**Ginger lethal concentration-50 (LC$_{50}$):**

The result of LC$_{50}$ of ginger for *C. gariepinus* after 96 hours appeared to be 192 mg/L (Table 2).

**Table 2: Conducting of ginger LC$_{50}$ for Clarias gariepinus after 96 hours.**

<table>
<thead>
<tr>
<th>Exposure dose (mg/L)</th>
<th>Fish Numbers/groups</th>
<th>No. of dead Fish</th>
<th>A</th>
<th>B</th>
<th>AXB</th>
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<tr>
<td>20</td>
<td>0</td>
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<td>0</td>
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<td>60</td>
<td>1</td>
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<td>80</td>
<td>2</td>
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<td>30.0</td>
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<tr>
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<td>10</td>
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<td></td>
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<td>120</td>
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<td>4</td>
<td>80.0</td>
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<td>10</td>
<td>9.5</td>
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<td>Σ</td>
<td>100</td>
<td>800</td>
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</tbody>
</table>

LC$_{50} = 192$ mg/L.

(A) is the mean of dead fish between two successive doses.

(B) is dose differences between two successive doses.

**Efficacy of ginger bath on the ectoparasites infesting C. gariepinus:**

The results of ginger efficacy on the infested *C. gariepinus* with mixed infestation of *Trichodina*, *Epistylis* and *Gyrodactylus* spp. indicated that long exposure of infested fish for three days (long bath) to ginger at concentration 10 mg/L was insufficient to eradicate the protozoa and monogenea while ginger concentration of 20 mg/L was sufficient to eradicate the protozoa (*Trichodina* and *Epistylis* spp.) but can not eradicate the monogenean (*Gyrodactylus* spp.) (Table 3). Overdose of ginger caused ulceration and erosion in the skin and fins of catfish (Fig. 6).
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Average was 15%. This result was lower than that recorded by El-Tantawy and Younis (2003) in Oreochromis niloticus (25%).

The average of monogenean Gyrodactylus spp. Fig. (4, 5) was 30% among infested C. gariepinus. This result was lower than that obtained by (Endraws, 2001). Also, a higher average (100%) was recorded by Paperna (1996) in the Nile fish. These variations in results might be attributed to the inhibitive quality of physical (Depth, current, temperature) and chemical (Oxygen, salinities) factors of the environment and fish species.

In the present study, the intensity of the ectoparasitic protozoa Trichodina, Epistylis and the monogenean Gyrodactylus spp. were 8-10, 15–18 and 6-8 parasites per field, respectively (Table 1). These numbers of ectoparasites per fish were considered a moderate infestation in O. niloticus as mentioned by El-Tantawy (2003). It may be attributed to that, in wild, catfish have many species of parasites with small numbers (FAO, 2007).

In Table (1), the total average of ectoparasites was 60%. This result was lower than that of (El-Tantawy, 2001) (100% for protozoa and 76% for monogenea) in Oreochromis niloticus and higher than that recorded by Diab et al. (2006) as (44.4%) among cultured C. gariepinus in Abassa. In addition, the average of the Trichodina spp. Fig. (2) in the infested catfish was 20% which was lower than the finding of Hanna (2001) who recorded Trichodina spp. from gills of common carp. With concern to Epistylis spp. Fig. (3), its average was 15%. This result was lower than that recorded by El-Tantawy and Younis (2003) in Oreochromis niloticus (25%).

In respect to the undesirable side effects of the chemical treatment of parasitic diseases and its public health hazards, the herbal treatment is taken place to relieve the harmful side effects of chemicals. So, in this study, the ginger was screened for its efficacy in treating C. gariepinus infested with a mixed infestation of Trichodina, Epistylis and Gyrodactylus spp. To avoid overdosing of ginger and to determine an effective control dose for ectoparasites, a bioassay of ginger LC$_{50}$ after 96 hours was conducted (Table 2). The value of 192mg/L ginger after 96 hours appeared to be its lethal concentration 50 for the exposed fish. So, the safe ginger dose must be 19.2mg/L which is 10 times

Discussion

All ectoparasites appear to be an important pathological stressor affecting fish populations in the water stratum. In the present study, the gross clinical appearance of infested C. gariepinus with ectoparasitic protozoa (Trichodina and Epistylis spp.) and the monogenean Gyrodactylus spp. demonstrated pale skin, excessive mucus secretion and skin erosion with local ulcerative lesions (Fig. 1a&b). These results were similar to that recorded by (Kuperman and Matey, 2000 and El-Tantawy, 2001).

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The taxonomy of the isolated Trichodina, Epistylis and Gyrodactylus spp. was typical to that reported by Paperna (1996), Huh et al. (2005) and Rotkiewicz (2006) Moreover, their morphological identification was similar to the description of El-khatib (1993), Lynn (2002) and Lome (2006).

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lower than such value as recommended by US Environmental protection Agency (Boyd, 1998) but in this study, the safe and effective dose of ginger for the exposed catfish was 20mg/L which was approximately near the calculated safe dose 19.2mg/L. The higher ginger doses than 20 mg/L caused various ulcerations and bloody erosions in the skin and fins of the treated fish (Fig. 6).

The efficacy of herbal treatment with ginger (long bath) on ectoparasites of catfish (Table 3) determined that the suitable dose of ginger for eradication of ectoparasitic protozoa Trichodina and Epistylis was 20mg/L for three successive days. On the other hand, neither the doses of 10mg/L nor 20mg/L of ginger was sufficient to treat the monogenean Gyrodactylus spp. That dose of ginger used as protozoacid was in agreement with Boyd (1998) who recorded that the safe level recommended by US Environmental protection Agency are 10 to 100 times lower than the value of 96 hrs LC 50. The pungent principles (including the volatile oil gingerol) are the most medicinally potent. It was performed to fish by Colorni (1987).

Concerning the impact of ginger on the water quality parameters, data shown in Table (4) revealed that the dissolved oxygen (5.6-6.7 ppm) was within its permissible limit (5-15 ppm) (Boyd, 1998). The pH values were approximately in the neutral range (6.92-7.2) in both pre and during treatment which is considered the ideal pH value for fish aquaculture operation (Noga, 2000). On the other hand, the recorded value of total ammonia (9-14.2 ppm) was much higher than its permissible limit (<0.1 ppm) (Boyd, 1998). These results might be attributed to the carnivorous behavior of catfish species that metabolize more protein in their food and consequently more ammonia released in water as an end product of metabolism. However, the recovered level of total ammonia is unaffected in the presence of the recorded neutral pH value, meanwhile, in the presence of alkaline pH, the more toxic free ammonia (NH3) will increase in water (FAO, 2007). In addition, this

<table>
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<tr>
<th>Fish groups</th>
<th>Ginger doses mg/l</th>
<th>Period of treatment/day</th>
<th>Trichodina, spp. 8/ field (10X)</th>
<th>Epistylis, spp. 18/ field (10X)</th>
<th>Gyrodactylus, spp. 6/ field (10X)</th>
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<td>1st group (15)</td>
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<td>8</td>
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Table 3: Efficacy of ginger bath on ectoparasites infesting C. gariepinus.

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<td>2nd day</td>
</tr>
<tr>
<td>1-Dissolved oxygen (PPM)</td>
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<td>6.4</td>
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<tr>
<td>2- pH</td>
<td>7.1</td>
<td>6.92</td>
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<tr>
<td>3-Total ammonia (PPM)</td>
<td>14.2</td>
<td>9</td>
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Table 4: Estimation of some water parameters pre and during ginger treatment (20 mg/L).
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study revealed that the values of total ammonia during applying ginger for three days (9, 10.1 and 11.2 ppm, respectively) were lower than its value before treatment (14.2 ppm), these results proved the significant usage of ginger to improve the water quality.

Conclusion

Ginger has a good effect on treating ectoparasitic protozoa *Trichodina* and *Epistylis* spp. at dose 20mg/L. It also improves the water quality for aquaculture operation with absence of undesirable side effects in comparison to other chemicals used in this trend.

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