

**Original Article**

**Evaluation of Biochar as Abio filter for Application in Commercial Fish Culture System**

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**Abstract**

Biochar is commonly used for absorption of heavy metals in the soils and water and so decreasing its level in fish meat with improvement of growth performance, productivity and hatchability as well as immunity of the fish against different fish diseases. The present work was conducted a fish farm is Metobase area in Kafr El-Sheikh governorate during season 2014. The period extended from January to May at the department of fish production Fac. of Agric. Saba Basha, Alexandria University. The biochar was prepared by burning of the rice bran at 300 °C using circular cylinder rotated by anaerobic motors. In our investigation, a total number of 162 broodstock of *Oreochromis niloticus* fish that were collected from private fish farm at Kafr El-Sheikh governorate, Egypt. The enhancement of aeration of aquaculture occurred by air blowers and agriculture machine, and the water column of ponds in both was about 1.5m. The aim of this study is to investigate the importance of biochar in growth performance and production efficiency of Nile tilapia as well as protective effects of heavy metal pollution in the aquaculture environment, product of fish and finally the histopathological alterations. Our results concluded that, the addition of biochar to the fish production ponds decrease the level of heavy metals and reduced its accumulation in fish organs especially the saw dust biochar, followed by rice husk biochar.

**Key words:** *Biochar, Oreochromis niloticus, Pollution, Heavy metals, Histopathology*

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## Introduction

Biochar is a fine-grained charcoal high in organic carbon and largely resistant to decomposition. It is produced from pyrolysis of plant and waste feedstocks. It's commonly used as a filter for the water especially drinking water or fish farm water. As a soil amendment, biochar creates a recalcitrant soil carbon pool that is carbon-negative, serving as a net withdrawal of atmospheric carbon dioxide stored in highly recalcitrant soil carbon stocks. The enhanced nutrient retention capacity of biochar-amended soil not only reduces the total fertilizer requirements but also the climate and environmental impact of croplands. Char-amended soils have shown 50 - 80 percent reductions in nitrous oxide emissions and reduced runoff of phosphorus into surface waters and leaching of nitrogen into groundwater. As a soil amendment, biochar significantly increases the efficiency of and reduces the need for traditional chemical fertilizers, while greatly enhancing crop yields. Renewable oils and gases co-produced in the pyrolysis process can be used as fuel or fuel feed-stocks. Biochar thus offers promise for its soil productivity and climate benefits (Major et al., 2010; Vaccari et al., 2011; Reverchon et al., 2014; Zhang and Sun, 2014). Using biochar as a soil fertilizers improved the plant growth, soil fertility with reduction of the heavy metal pollution in the agriculture water with and its usage in aquaculture ponds improved the fish growth than the non-fertilized soil or aquaculture ponds with biochar (Schmidt, 2014). Yang et al. (2014) suggested that using of a low-cost *Alternanthera philoxeroides* biochar (APB) adsorbent for heavy metals contaminated water treatment may have great ecological and environmental significance as it reduces the level of heavy metals such as lead, mercury, copper and other harmful heavy metals. Lim et al. (2014) observed that the using of biochar in aquaculture may potentially alter nitrous oxide (N<sub>2</sub>O) emission by affecting ammonia-oxidizing

and nitrification bacteria, which is determined by the application rate of biochar in soil and water. The means by which biochar improves production response can be attributed to direct effects via biochar-supplied nutrients (Silber *et al.*, 2010), and to several other indirect effects, including: increased nutrient retention; improvements in soil pH; increased soil action exchange capacity (Yamato *et al.*, 2006); effects on Phosphorus and Sulfur transformations and turnover; neutralization of phytotoxic compounds in the soil (Wardle *et al.*, 1998). Biochar improved soil physical properties including water retention; promotion of mycorrhizal fungi (Warnock *et al.*, 2007); and alteration of soil microbial populations and functions (Graber *et al.*, 2010 and Kolton *et al.*, 2011). Given that the biochar-soil-plant-water environment is highly complex, it is difficult to isolate those factors which actually play an instrumental role in 'The Biochar Effect'. To reduce the number of potential factors involved, (Pietikainen *et al.*, 2000), tested whether biochar addition could impact plant growth when nutritional and soil physical aspects of biochar amendment were eliminated, by using a nutrient-poor, wood-derived biochar, under an optimal fertilization and irrigation regime in a greenhouse(Graber et al., 2010). They reported an increase in several plant growth parameters for both plants by using biochar-treatment. Environmental pollution represents a major problem in the world, especially in the less developed countries. Egypt is one of those countries which suffer from biosphere pollution (air, soil and water). Agricultural, industrial and sewage wastes, which are discharged into the River Nile, canals and other water resources causing ecological changes in water. Pollutants including heavy metals, pesticides, hydrocarbons that present in polluted water way be accumulated in fish tissues and organs causing severe healthy problems to the consumers.

The aim of this study is to investigate the importance of biochar in prevention of

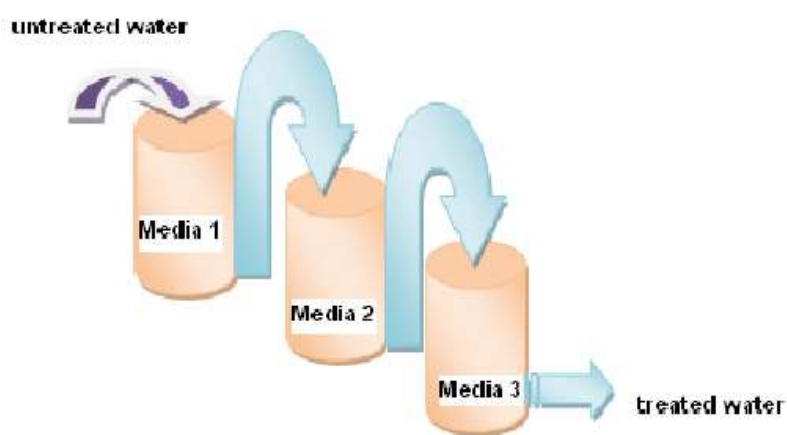
heavy metal pollution in the aquaculture environment of either water, product of fish or the decreasing the level of histopathological changes that observed in the fish organs due to heavy metal pollutions. Through study of the level of heavy metals in water, fish products and the histopathological changes resulted in heavy metal pollutions and biochar addition to the fish ponds.

### Materials and methods

Experimental broodstock trial was carried out in fish farm in Metobase area Kafr El-Sheikh Governorate in triplicate during 2014. Adult 162 broodstock *Oreochromis niloticus* with an average body weight  $482.8 \pm 2$  g/ fish were used to evaluate Biochar from different compound as a biofilters for application in commercial fish culture system. Fish were fed on a commercial fish diet containing 28.4% crude protein. The diet was daily provided with specific feeding ratio of according to body weight and growth stages of fish as described by Eurell *et al.* (1978) as follow. Feeding ratio

for fish about was 4 % of their life body weight.

The biochar filters media of rice husk (RH), saw dust (SD), clay brick dust (CBD) and sand (S) used in this study were collected from the local market. The collected materials were extensively washed with tap water to remove soil and dust. Dry RH and SD were crushed and sieved by 0.5-mm polypropylene sieve for preparing the biochar materials. A pilot-scale diesel pyrolyzer designed at the central Laboratory for Agriculture Climate, with a maximum capacity to process 15 kg of raw material per batch was used to produce biochar. Carbonization temperatures used for both types of residual biomass were 350°C. The temperature was increased at a rate of 5 °C/min until the specific temperature was reached and maintained for 1 h. After that, a cooling down procedure until room temperature was carried out. Biochar filter materials and raw materials were placed in pocket's in three steps media 1, 2 and 3 according to Table 1 and Fig1.



This drawing to illustrate one sample (Group) of the filter used in this study

**Fig 1.** Illustrate sample (Group) of the filter used in this study

**Table 1.** Design of the experiment

Groups	Treatment		
	Media 1	Media 2	Media 3
Group1	Sand	Raw rice husk	Raw rice husk
Group2	Sand	Break dust	Biochar saw dust
Group3	Sand	Biochar saw dust	Biochar saw dust
Group4	Sand	Biochar rice husk	Biochar rice husk
Group5	Sand	Break dust	Biochar saw dust
Group6	Sand	Break dust	Biochar rice husk
Group7	Sand	Biochar rice husk	Biochar saw dust
Group8	Sand	Break dust	raw rice husk
Group9	Control		

Water samples were collected before treatment and after treatment from different groups to determine the water chemical characteristics (pH, NH<sub>3</sub>, Dissolved Oxygen and EC) and heavy metals (Pb, Cd, Zn, Cu, Hg, Fe, Ni and Si). Dissolved oxygen (mgL<sup>-1</sup>) meter of the model: OxyGuard Handy MK II was used in measuring dissolved oxygen and temperature. PH was measured using pH meter (model: Hanna Instrument model No. HI 8915 ATC) while salinity was measured using salinometer, model: New S-100 for each of the parameters. The method for analysis of the heavy metals in the water was carried out according to AOAC (1996) and in the fish tissues according to Clesceri (1998) that was carried out using Atomic Absorption Spectrophotometer. Atomic Absorption Spectrophotometer (Model Thermo Electron Corporation, S. Series AA Spectrometer with Gravities furnace, UK,) instrument was used to detect the heavy metals. The concentrations of heavy metals were expressed as mgL<sup>-1</sup> for water and µg g<sup>-1</sup> for fish tissue dry weight. Histopathological examination was made according to the methods implied by (Roberts *et al.*, 1993) and the sections were stained with H & E stain and examined microscopically. Statistical analysis: The data of bacteriological examinations and heavy metal concentrations were statistically analyzed using ANOVA tests according to (SAS, 2004) for detecting the differences in metal concentration among treatments.

## Results and discussion

Water quality is very important factor in the fish hatchery, especially out of heavy metals. Biochar is able by its mechanical treatment of wastewater (i.e. sieves, rakes, followed by a decantation process) followed by a biological cleaning process (i.e. Sewage fields, filtration, trickling filter, submerged bodies, sludge process, nitrate system, decay system) to clear the water out of pollutants (Imhoff *et al.*, 2009). In addition to, shows that organic substances can be removed through absorption with activated carbon. Pollutants also as heavy metals, pesticides and substances from anthropogenic origin remain a huge issue in water can be removed by biochar (Jonathan, 2013). Iwamoto (2012), observed that, the addition of biochar (Iwamoto Biochar for Aquaculture) to the fish and shrimp culture ponds improve the immunity of the fish and shrimp, body weight gain and body weight than the shrimp grown in the ponds without any biochar additives. It worthy to be noted that the increase of survival rate is closely related to the immunity of the fis. Table 2, and figure 2, summarized the weight (WT), Total production (Total PR), Production/female (PPF) and Production/gram (PPG) in broodstock of tilapia in different groups of experiment. The data revealed that the WT were

**Table 2.** Body weight, total production, production/female and production/gram in broodstock of tilapia in different groups of experiment

Groups	WT	TOTALPR	PPF	PPG	SR
	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE
G1	482.50 $\pm$ 1.44 <sup>a</sup>	6234.00 $\pm$ 176.89 <sup>bc</sup>	779.25 $\pm$ 22.11 <sup>bc</sup>	1.61 $\pm$ 0.05 <sup>bc</sup>	92.13 $\pm$ 0.52 <sup>b</sup>
G2	481.67 $\pm$ 1.10 <sup>a</sup>	6474.00 $\pm$ 14.47 <sup>b</sup>	809.25 $\pm$ 1.81 <sup>b</sup>	1.68 $\pm$ 0.00 <sup>b</sup>	91.13 $\pm$ 0.70 <sup>bcd</sup>
G3	482.08 $\pm$ 0.75 <sup>a</sup>	5800.00 $\pm$ 230.94 <sup>bc</sup>	725.00 $\pm$ 28.87 <sup>bc</sup>	1.50 $\pm$ 0.06 <sup>cd</sup>	90.17 $\pm$ 0.73 <sup>cd</sup>
G4	482.50 $\pm$ 0.72 <sup>a</sup>	7192.33 $\pm$ 137.70 <sup>a</sup>	899.00 $\pm$ 17.20 <sup>a</sup>	1.86 $\pm$ 0.04 <sup>a</sup>	94.53 $\pm$ 0.43 <sup>a</sup>
G5	482.50 $\pm$ 0.72 <sup>a</sup>	5920.33 $\pm$ 119.35 <sup>bc</sup>	740.02 $\pm$ 14.92 <sup>bc</sup>	1.53 $\pm$ 0.03 <sup>bcd</sup>	91.83 $\pm$ 0.26 <sup>bc</sup>
G6	482.92 $\pm$ 0.75 <sup>a</sup>	5812.00 $\pm$ 299.74 <sup>bc</sup>	726.50 $\pm$ 37.47 <sup>bc</sup>	1.50 $\pm$ 0.08 <sup>cd</sup>	90.60 $\pm$ 0.52 <sup>bcd</sup>
G7	482.71 $\pm$ 0.55 <sup>a</sup>	6132.00 $\pm$ 227.60 <sup>bc</sup>	728.97 $\pm$ 12.37 <sup>bc</sup>	1.51 $\pm$ 0.03 <sup>cd</sup>	91.97 $\pm$ 0.34 <sup>b</sup>
G8	482.29 $\pm$ 0.91 <sup>a</sup>	5656.33 $\pm$ 345.88 <sup>c</sup>	707.00 $\pm$ 43.23 <sup>c</sup>	1.47 $\pm$ 0.09 <sup>cd</sup>	90.17 $\pm$ 0.64 <sup>cd</sup>
G9	482.29 $\pm$ 0.55 <sup>a</sup>	5681.00 $\pm$ 226.92 <sup>c</sup>	710.07 $\pm$ 28.35 <sup>c</sup>	1.41 $\pm$ 0.03 <sup>d</sup>	89.87 $\pm$ 0.28 <sup>d</sup>

**Table 3.** Heavy metal (Cu, Zn, cd, Pb, Hg and Fe) concentrations (mg/litre) in water samples in different groups of experiment

Groups	Cu	Zn	Cd	Pb	Hg	Fe
	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE
G1	0.407 $\pm$ 0.035 <sup>b</sup>	2.207 $\pm$ 0.063 <sup>ab</sup>	0.032 $\pm$ 0.005 <sup>b</sup>	0.058 $\pm$ 0.007 <sup>b</sup>	0.002 $\pm$ 0.000 <sup>b</sup>	1.219 $\pm$ 0.011 <sup>c</sup>
G2	0.303 $\pm$ 0.012 <sup>c</sup>	2.370 $\pm$ 0.112 <sup>a</sup>	0.037 $\pm$ 0.002 <sup>b</sup>	0.048 $\pm$ 0.005 <sup>b</sup>	0.003 $\pm$ 0.000 <sup>ab</sup>	1.289 $\pm$ 0.035 <sup>b</sup>
G3	0.024 $\pm$ 0.004 <sup>d</sup>	2.321 $\pm$ 0.083 <sup>a</sup>	0.003 $\pm$ 0.001 <sup>c</sup>	0.014 $\pm$ 0.002 <sup>c</sup>	0.001 $\pm$ 0.000 <sup>c</sup>	0.688 $\pm$ 0.028 <sup>d</sup>
G4	0.019 $\pm$ 0.002 <sup>d</sup>	2.020 $\pm$ 0.002 <sup>b</sup>	0.002 $\pm$ 0.000 <sup>c</sup>	0.017 $\pm$ 0.001 <sup>c</sup>	0.001 $\pm$ 0.000 <sup>c</sup>	0.518 $\pm$ 0.030 <sup>e</sup>
G5	0.277 $\pm$ 0.018 <sup>c</sup>	2.041 $\pm$ 0.012 <sup>b</sup>	0.005 $\pm$ 0.001 <sup>c</sup>	0.053 $\pm$ 0.006 <sup>b</sup>	0.003 $\pm$ 0.000 <sup>b</sup>	1.370 $\pm$ 0.059 <sup>b</sup>
G6	0.035 $\pm$ 0.012 <sup>d</sup>	2.190 $\pm$ 0.022 <sup>ab</sup>	0.002 $\pm$ 0.000	0.014 $\pm$ 0.003 <sup>c</sup>	0.000 $\pm$ 0.000 <sup>c</sup>	0.448 $\pm$ 0.006 <sup>e</sup>
G7	0.320 $\pm$ 0.021 <sup>c</sup>	2.249 $\pm$ 0.032 <sup>ab</sup>	0.027 $\pm$ 0.008 <sup>b</sup>	0.045 $\pm$ 0.004 <sup>b</sup>	0.002 $\pm$ 0.000 <sup>b</sup>	1.354 $\pm$ 0.035 <sup>b</sup>
G8	0.020 $\pm$ 0.008 <sup>d</sup>	2.242 $\pm$ 0.030 <sup>ab</sup>	0.001 $\pm$ 0.000 <sup>c</sup>	0.012 $\pm$ 0.001 <sup>c</sup>	0.000 $\pm$ 0.000 <sup>c</sup>	0.381 $\pm$ 0.012 <sup>f</sup>
G9	0.600 $\pm$ 0.040 <sup>a</sup>	2.377 $\pm$ 0.148 <sup>a</sup>	0.074 $\pm$ 0.006 <sup>a</sup>	0.094 $\pm$ 0.004 <sup>a</sup>	0.004 $\pm$ 0.000 <sup>a</sup>	1.568 $\pm$ 0.040 <sup>a</sup>

highest in Groups 6 and 7, while TOTALPR in Groups 1, 2, 4 and 7, the

PPF in Groups 1, 2 and 7 and the PPG were highest in Groups 1, 2 and 7. On the

other hand the other groups were equals in the values. The results of reproduction in broodstock of tilapia in presence of biochar is strongly related to absence of stress and good quality of water during hatchery which directly improvement of spawning. The different authors reported the biochar has many potential benefits on aquaculture process (Lehmann et al., 2011; Gascó *et al.*, 2012 and Paz Ferreiro *et al.*, 2014;). Also, biochar addition to the water will cause alterations in water quality (Paz-Ferreiro and Fu, 2014) with the potential to increase agricultural yields (Jeffery *et al.*, 2011; Liu *et al.*, 2013). In addition to enhance biochar' ability to immobilize heavy metals (Lima *et al.*, 2014). Interaction between biochar and heavy metal it is necessary to note that biochars act on the bioavailable fraction of soil heavy metals and that they can reduce

Tables 3 and 4 illustrated the heavy metals in the water in different groups of experiment. The data revealed that the heavy metals were lowest in the Groups 3, 4, 5, 6 and 8. On the same manner the other groups the same heavy metals were higher than the treated groups but in the same time is lower than the permissible limits. Tables (3 and 4) cleared the significant differences of the levels of different heavy metals in water among different treated groups ( $P < 0.01$ ). The results cleared that the zinc level showed a higher level in control group (2.38) and the lower level observed biochar saw dust (2.02), the level of copper showed a higher level in control group (0.60) and the lower level observed in group 8 that treated with sand + break dust + raw rice husk. (0.02). The iron level showed a higher level in control group (1.57) and the lower level observed in group 6 that treated with sand + biochar rice husk + biochar saw dust. (0.16). The Nickel level showed a higher level in control group (2.26) and the lower level observed in group 8 that treated with sand + break dust + raw rice husk. The

also their hatchability (Beesley and Marmiroli, 2011 and Lu *et al.*, 2012). Fry survival rate had been increased, may be due to the improvement induced by using of biochar. Alexander and Stewart, 2001 and An *et al.*, 2009 reported that *Flavobacterium* isolates have been shown to have biocontrol capabilities and to elicit resistance response of plants to different diseases during used of biochar. Graber *et al.* (2010) also, isolated a number of bacteria with identity to known biocontrol agents, induced resistance agents and growth promoters (15 out of 20 total isolates) from the root zone of biochar-amended pepper plants where promotion of plant growth and induction of systemic resistance against Fungal foliar diseases occurred simultaneously (Elad *et al.*, 2010 and Graber *et al.*, 2010).

mercury level showed a higher level in control group (0.0043) and the lower level observed in group 8 that treated with sand + break dust + raw rice husk. (0.0004) and group 6 that treated with Biochar rice husk + Biochar saw dust (0.0004). The cadmium level showed a higher level in group A (0.074) and the lower level observed in group 8 (0.0013) that treated with Sand + Break dust + raw rice husk. The zinc level showed a higher level in control group (0.09) and the lower level observed in the group (3) that treated with Sand + Biochar saw dust + Biochar saw dust, group (4) that treated with Sand + Break dust + Biochar saw dust, group (6) and group (8) as its level reached to (0.01). The silicon level showed a higher level in group (2) (2.22) and the lower level observed in group (8) that treated with Sand + Break dust + raw rice husks its level reached to (0.14). The arsenic level showed a higher level in control group (0.0013), group 1 (0.0003), group (3) that treated with Sand + Biochar saw dust + Biochar saw dust as its level reached to (0.0003) and the lower level reached to (0)

**Table 4.** Heavy metal (Ni, Si and Chromium) concentrations (mg/litre) in water samples in different groups of experiment

Groups	Ni	Si	chromium
	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE
G1	1.330 $\pm$ 0.059c	1.947 $\pm$ 0.176b	0.069 $\pm$ 0.008a
G2	1.783 $\pm$ 0.063b	2.223 $\pm$ 0.167a	0.061 $\pm$ 0.004a
G3	0.794 $\pm$ 0.070e	0.233 $\pm$ 0.065d	0.009 $\pm$ 0.002c
G4	0.497 $\pm$ 0.041f	0.230 $\pm$ 0.023d	0.012 $\pm$ 0.001bc
G5	1.197 $\pm$ 0.068cd	1.397 $\pm$ 0.055c	0.066 $\pm$ 0.005a
G6	0.161 $\pm$ 0.045g	0.260 $\pm$ 0.025d	0.007 $\pm$ 0.000c
G7	1.073 $\pm$ 0.143d	1.733 $\pm$ 0.046b	0.049 $\pm$ 0.005a
G8	0.177 $\pm$ 0.030g	0.147 $\pm$ 0.012d	0.008 $\pm$ 0.001c
G9	2.260 $\pm$ 0.137a	1.220 $\pm$ 0.067c	0.040 $\pm$ 0.027ab

**Table 5.** Heavy metals (Cu, Zn, Cd, Pb, Hg and Fe) concentrations ( $\mu$ g/g dry wt.) in liver of *Oreochromis niloticus* in different groups of experiment

Groups	Cu	Zn	Cd	Pb	Hg	Fe
	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE	Mean $\pm$ SE
G1	33.93 $\pm$ 0.84a	176.63 $\pm$ 8.77a	0.23 $\pm$ 0.01a	0.45 $\pm$ 0.02a	1.60 $\pm$ 0.10a	138.42 $\pm$ 7.47a
G2	29.01 $\pm$ 1.12b	140.76 $\pm$ 1.86b	0.25 $\pm$ 0.01a	0.46 $\pm$ 0.02a	1.20 $\pm$ 0.04bc	118.44 $\pm$ 1.35b
G3	26.17 $\pm$ 1.79b	122.58 $\pm$ 1.58c	0.22 $\pm$ 0.00a	0.41 $\pm$ 0.01a	1.42 $\pm$ 0.06ab	102.82 $\pm$ 2.22c
G4	3.93 $\pm$ 1.24f	62.63 $\pm$ 1.62h	0.13 $\pm$ 0.02b	0.23 $\pm$ 0.01c	0.33 $\pm$ 0.20d	38.17 $\pm$ 3.15f
G5	9.19 $\pm$ 0.31de	73.61 $\pm$ 4.13gh	0.12 $\pm$ 0.01bc	0.29 $\pm$ 0.02b	0.33 $\pm$ 0.20d	56.73 $\pm$ 3.39e
G6	21.05 $\pm$ 0.94c	108.70 $\pm$ 2.14d	0.23 $\pm$ 0.00a	0.43 $\pm$ 0.01a	1.07 $\pm$ 0.03c	115.58 $\pm$ 5.03bc
G7	20.55 $\pm$ 0.95c	101.17 $\pm$ 4.50de	0.22 $\pm$ 0.01a	0.45 $\pm$ 0.02a	1.11 $\pm$ 0.06bc	110.45 $\pm$ 6.08bc
G8	7.77 $\pm$ 0.81e	85.34 $\pm$ 4.08fg	0.10 $\pm$ 0.02bc	0.22 $\pm$ 0.00c	0.08 $\pm$ 0.03d	66.72 $\pm$ 2.89de
G9	11.91 $\pm$ 1.16d	93.88 $\pm$ 2.55ef	0.09 $\pm$ 0.02c	0.26 $\pm$ 0.02bc	0.16 $\pm$ 0.06d	74.77 $\pm$ 6.62d

that observed in the groups (3), (4) that treated with Sand + Break dust + Biochar saw dust, (6) that treated with Sand + Biochar rice husk + Biochar saw dust, group (7) that treated with Sand + Break dust + Biochar rice husk and group (8) that treated with Sand + Break dust + raw rice husk. The chromium level showed a higher level that, observed in control group (0.04), and group (7) that treated with Sand + Break dust + Biochar rice husk and its level reached to (0.04) . While, the lower level observed in group (8) that treated with Sand + Break dust + raw rice husks its level reached to (0.007). The manganese level showed a higher level in control group (0.07) and the lower level that observed in group 3 and 4 as its level reached to (0.01). Our results cleared that, the biochar addition to the fish ponds decreasing the level of heavy metals in water due to the biochar absorbed the heavy metals in the water with its reduction level than the other treatment especially the biochar dust than the other types of biochar. Our results agreed with those of (Silber *et al.*, 2010 and Yang *et al.*, 2014), where they observed that, the addition of biochar to fish ponds reduce the level of heavy metals in fish production ponds.

Table, 5 illustrated the heavy metals in the liver in different groups of experiment. The data revealed that the heavy metals were lowest in the Groups 3, 4, 5, 6 and 8. On the same manner the other groups the same heavy metals were higher than the treated groups but in the same time is lower than the permissible limits. Table (14) cleared the significant differences of the level of heavy metals in liver among different treatments among examined fish product samples. Zinc level showed a higher level in biochar rice husk (85.10) followed by raw rice husk (73.66) and the lower level of zinc observed in the group treated with Biochar saw dust (0.34) and red break dust (3.15) . The copper level showed a high level in raw rice husk (85.33) and biochar rice husk (46.33) and the lower level observed in biochar saw dust (4.41) and red break (0).

The iron level showed a higher level in raw rice husk (139.85) followed by Biochar rice husk (96.59) and the lower level observed in biochar saw dust (0.54). The nickel level show a high level in raw rice husk (1.93) and the lower level of nickel observed in the biochar rice husk (0.65). The mercury level not recorded among the all treatments and its level reached to (0). The cadmium level not recorded in all treatments and its level reached to (0) but in raw rice husk reached to (0.06). The lead level not recorded in all treated groups except in red break treated groups (18.05). The silicon level showed a higher level in sand treated groups (671.17) and the lower level observed in biochar saw dust (78.25). The Arsenic level not recorded among the all treatments and its level reached to (0). The chromium level in all groups showed a level of (0) except in the groups treated with raw rice husk (44.19) and Biochar rice husk (41.07). The manganese level showed a higher level in Biochar saw dust (720.12) and the lower manganese level observed in sand treated group (173.39). Our results concluded that, the biochar increase the level of secretion of heavy metals in fish products than the other treatments especially the rice husk biochar and saw dust biochar. This results attributed to the biochar activated the fish viability and immunity also the biochar contain a higher level of heavy metals especially arsenic, iron and nickel. Our results agreed with those of Fang *et al.* (2014) and Zhang *et al.* (2012), where they reported that the biochar improve the secretion of heavy metals in fish products due to improving the fish viability and digestibility of the foods.

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